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LIMITED IMPACT OF ROSELLE FLOWER EXTRACT ON PROTEIN CARBONYL LEVELS AFTER PHYSICAL EXERCISE IN HEALTHY MEN

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Key words: roselle flower extract, oxidative stress, protein carbonyl, antioxidants, exercise-induced muscle damage **Ключові слова:** екстракт квітів розелли, окиснювальний стрес, карбоніл білка, антиоксиданти, пошкодження м'язів, спричинене фізичним навантаженням

Abstract. Limited impact of roselle flower extract on protein carbonyl levels after physical exercise in healthy men. Ayubi N., Rusdiawan A., Hartoto S., Wibawa J.C., Rizki A.Z., Afandi A., Wardani A.P.S., Halip M.F., Jr P.B.D. Exercise-induced oxidative stress can lead to muscle damage, with protein carbonyls serving as a key biomarker of oxidative modification in proteins. Roselle flower extract contains polyphenols with potential antioxidant properties that may reduce oxidative damage after physical exercise. This study investigated whether post-exercise supplementation of roselle flower extract reduces protein carbonyl levels. A randomized controlled trial with a parallel group pre-test/post-test design was conducted, comparing a placebo group with a treatment group receiving 500 mg of roselle flower extract. The extract did not result in a statistically significant reduction in protein carbonyl levels (p>0.05). However, a non-significant decreasing trend was observed, suggesting that future studies should explore larger sample sizes and longer intervention periods to assess potential cumulative effects. Further studies should explore the effects of higher doses of roselle flower extract (e.g., >500 mg/day) and chronic supplementation over several weeks to determine its long-term impact on markers of oxidative stress.

Реферат. Обмежений вплив екстракту квітів розелли на рівень карбонілів білка після фізичних вправ у здорових чоловіків. Аюбі Н., Русдіаван А., Хартото С., Вібава Дж.К., Різкі А.З., Афанді А., Вардані А.П.С., Халіп М.Ф., Дж Р. П.Б.Д. Окиснювальний стрес, спричинений фізичними вправами, може призвести до пошкодження м'язів, при цьому білкові карбоніли служать ключовим біомаркером окисної модифікації білків. Екстракт квітів розелли містить поліфеноли з потенційними антиоксидантними властивостями, які можуть зменишти окисне пошкодження після фізичних вправ. У цьому дослідженні було з'ясовано, чи споживання екстракту квітів розелли після тренування знижує рівень карбонілів білка. Було проведено рандомізоване контрольоване дослідження з паралельною групою до/посттестового періоду, у якому порівнювались група плацебо з групою лікування, яка отримувала 500 мг екстракту квітів розелли. Екстракт не призводив до статистично значущого зниження рівня карбонілу білка (p>0,05). Проте спостерігалася незначна тенденція до зниження, що свідчить про те, що в майбутніх дослідженнях слід вивчати більші розміри вибірки та триваліші періоди втручання для оцінки потенційних кумулятивних ефектів. У подальших дослідженнях має бути вивчено вплив високих доз екстракту квітів розелого прийому добавок протягом кількох тижнів, щоб визначити його довгостроковий вплив на маркери окисного стресу.

Reactive oxygen species (ROS) are continuously generated as by-products of cellular metabolism, especially in mitochondria. When ROS production exceeds the body's antioxidant defense mechanisms, oxidative stress occurs, leading to cellular damage [1]. Prolonged oxidative stress, if unbalanced by antioxidants, contributes to diseases like diabetes, hypertension, cardiovascular disease, cancer, and neurodegenerative disorders [2]. Exercise-induced ROS production is a double-edged sword. While excessive ROS can cause oxidative damage, moderate increases in ROS act as signaling molecules that enhance antioxidant defense mechanisms, mitochondrial biogenesis, and overall cellular adaptation to stress, contributing to long-term health benefits [3]. Depending on the length, level of intensity, and individual's level of physical fitness, exercise can have both beneficial and detrimental effects [4]. However, if the production of ROS is excessive, this will cause oxidative stress to develop and a change in the redox balance toward oxidation, if allowed to continue, will affect physiological mechanisms and disorders in the immune system [1]. Oxidative stress also adversely affects athlete performance and fatigue, which in turn leads to weakened immune function and increased susceptibility to infection [5].

The main endogenous sources of ROS in skeletal muscle include mitochondria, NADPH oxidase (NOX), and xanthine oxidase (XO) [6]. Xanthine oxidase (XO), present in muscle endothelium and cytosol, contributes to the generation of extracellular superoxide (O₂⁻) during metabolic stress and ischemiareperfusion conditions. While XO-derived ROS may play a role in redox signaling, excessive activity may contribute to oxidative damage rather than muscle force generation [7]. In addition, autooxidation of myoglobin or oxidation of hemoglobin to methemoglobin further contributes to oxidative stress in muscle by inducing the formation of peroxide [8]. It can be concluded that the mechanism of increased oxidative stress occurs during the formation of ATP in the mitochondria and then an increase in ROS during the respiration process which triggers muscle damage if antioxidant levels are not able to neutralize ROS [9].

One of the primary by products of free radicals breaking down unsaturated fatty acids is malondialdehyde (MDA), a biomarker of oxidative stress that is created by a class of free radicals known as hydroxyl radicals that induce lipid peroxidation [10]. Another biomarker of oxidative stress is protein carbonyls, which have the advantage of being relatively early in the formation and stability of protein carbonylation [11]. Protein carbonylation is one of the most widely used biomarkers of protein oxidation. Protein carbonylation occurs when ROS modify amino acid side chains, leading to irreversible structural and functional changes in proteins. Proteins with multiple oxidative pathways frequently experience loss of function and alterations in biological activity due to carbonylation, an irreversible deformation brought on by oxidative stress [12].

Protein carbonyl (PC) is considered a reliable biomarker of oxidative stress associated with cell damage, aging, and various age-related disorders [13]. Increased protein carbonyl levels are regarded as an early indicator of ROS exposure since the process is irreversible [14]. During strenuous physical training, data showed that athletes who contributed to the 50 km and 100 km ultramarathon races experienced a significant increase in protein carbonyls [15]. Extreme exertion, such ultra-marathon running, can increase oxygen use by 10-20 times, which will always produce reactive oxygen species (ROS) [1]. Numerous enzymatic and nonenzymatic antioxidants in biological systems can be elevated by exerciseinduced ROS, according to recent studies, despite early studies' unfavorable effects [16]. The body has an antioxidant defense system to fight against the oxidative stress that results. Exercise and diet can have an impact on antioxidants body's system consisting of enzymatic and non-enzymatic [17].

Roselle flowers are among the natural materials that possess the capacity to be used as anti-inflammatory and antioxidant substances. Hibiscus Sabdariffa L., commonly referred to as Roselle, is a potent antioxidant due to its polyphenols (anthocyanins, flavonoids, phenolic acids, tannins), polysaccharides, pectin, nonphenolic organic acids, carotenoids, caffeic acid, chlorogenic acid, ascorbic acid, and quercetin are examples of vitamins, minerals, and bioactive components [18]. Anthocyanin is one of the flavonoid polyphenols with potent antioxidant qualities [19]. Anthocyanins in Roselle exhibit strong antioxidant properties by scavenging superoxide anions and other ROS. In addition, they can enhance the activity of endogenous antioxidant enzymes, which contributes to the reduction of oxidative stress and inhibition of protein glycation [20]. Given that Rosella has proven antihyperlipidemic, antihyperglycemic, antioxidant, anti-inflammatory, antihypertensive, and antifibrosis qualities, it may also be investigated further as a drug to lower the risk factor for cardiovascular disease [21].

Roselle is known to have high antioxidant content. Roselle flower extract is believed to be an antioxidant that can reduce free radical levels. The potential of roselle flower extract to reduce exercise-induced oxidative stress by reducing protein carbonyl levels remains unclear, as few studies have directly examined this relationship. Therefore, this study aimed



to evaluate the effects of post-exercise Roselle flower extract supplementation on protein carbonyl biomarkers as indicators of oxidative stress. Therefore, the purpose of this study was to determine the effect of giving roselle flower extract after physical exercise on the oxidative stress response through protein carbonyl biomarkers.

MATERIALS AND METHODS OF RESEARCH

This experimental study used a before and after control group design. Subjects were selected using purposeful sampling based on inclusion and exclusion criteria. After selection, they were randomly assigned to one of two groups: K1 (placebo) or K2 (roselle flower extract, 500 mg). The 500 mg dose of roselle flower extract was chosen based on its safety profile established in human studies. Previous studies have shown that higher doses may provide stronger antioxidant effects, but for ethical considerations and consistency with existing literature, the 500 mg dose was chosen [18]. Supplementation was given in capsule form. Placebo is formulated using only empty capsules that are not given any contents.

Twenty men in good health took part in the study (Table 1 displays subject characteristics). To determine whether volunteers could meet the needs of this study, inclusion and exclusion criteria were established. Students with a normal Body Mass Index (BMI) between the ages of 20 and 25 years were included in the inclusion criteria. Participants were required to be non-regular exercisers, defined as engaging in structured physical activity less than three times per week for at least six months, as assessed via a pre-screening questionnaire. In addition, this study did not involve participants who were under 20 years of age or who had high blood pressure before engaging in physical activity. Additionally, if subjects were taking nonsteroidal anti-inflammatory drugs (NSAIDs), they were excluded. 20 research volunteers were divided into 2 groups, namely the physical exercise + roselle treatment group (n=10), and the control group with physical exercise + placebo (n=10).

Some of the instruments used in this study were blood pressure gauges, height gauges, weight gauges, data collection sheets, stationery, roselle flower extract, and placebo capsules.

The data collection process in this study involved several steps. The research subjects underwent a screening procedure before starting the study. Certain parameters that allowed the data to be included or excluded from the analysis formed the basis of this approach. In addition, they provided consent, agreeing to participate in the study. This research instrument complies with the principles of the Helsinki Declaration and has passed ethical review. The study subjects were randomly divided into two groups: the treatment group, which received roselle flower extract, and the placebo group. The treatment group consumed 500 mg of roselle flower extract, while the placebo group consumed empty capsules. The roselle flower extract was administered in capsule form.

Data collection was carried out for 1 day starting with collecting data on subject characteristics. The research subjects were prohibited from consuming anything before the implementation of the study. One day before the study, the research subjects were given directions to maintain a regular diet and rest pattern. After that, they were asked to warm up. All subjects performed a 2400-meter run at a self-selected moderate-intensity pace, with heart rate monitoring to ensure they remained within 50-70% of their agepredicted maximum heart rate (Hrmax = 220 - age). After the running test, the sample was directed to a shady room not far from the field to rest and wait for the next intervention. Venous blood samples were collected via antecubital venipuncture before and 60 minutes after supplementation. Blood was immediately centrifuged at 3000 rpm for 10 minutes at 4°C, and plasma was stored at -80°C until analysis to prevent oxidative degradation. Blood samples were collected 60 minutes post-intervention, based on previous studies showing that peak absorption of polyphenolic compounds from roselle flower extract occurs within this time frame [22, 23]. However, direct pharmacokinetic measurements were not performed in this study. The analysis used to determine protein carbonyl levels with using the Protein Carbonyl ELISA Kit was carried out at the Faculty of Medicine Laboratory, Brawijaya University Malang, Indonesia. Finally, the researchers analyzed the data and made a written report as a form of accountability (Fig. 1).

analysis was performed Statistical using SPSS 16.0 after data collection. Descriptive statistics was used to summarize the data, including means and standard errors for normally distributed variables and medians for non-normally distributed variables. Normality was assessed using the Shapiro-Wilk test. If data followed a normal distribution (p>0.05), a paired t-test was used to compare pre- and post-test values within groups. If the normality assumption was violated (p<0.05), the Wilcoxon signed-rank test was applied instead. Statistical significance was defined as p<0.05 for all analyses.

This study was conducted in accordance with the principles of the Declaration of Helsinki and has been approved by the Ethics Committee of the Malang Health Polytechnic (Approval Number DP.04.03/F.XXI.31/0486/2024). Written informed consent was obtained from all participants prior to enrollment.



Fig. 1. The CONSORT flowchart

RESULTS AND DISCUSSION

This section presents the data and provides information about the general characteristics of the participants in Table 1. These data allow us to better understand the characteristics of each group. Data are presented as mean \pm standard error. Independent

t-tests were performed to compare baseline characteristics between Group K1 and Group K2. The results showed no significant differences between the groups for age, height, weight, BMI, or blood pressure ($p \ge 0.05$ for all comparisons).

Table 1

Data	Group	Ν	x±SD	p-value		
Age (y)	K1	10	23.70±0.51	0.652		
	K2	10	23.30±0.77			
Height (cm)	K1	10	168.70±1.77	0.701		
	K2	10	170.20±2.27			
Weight (kg)	K1	10	62.20±2.62	0.722		
	K2	10	64.00±3.97			
BMI (kg/m ²)	K1	10	21.79±1.99	0.851		
	K2	10	21.99±1.06			
Systolic (mmHg)	K1	10	122.80±3.38	0.962		
	K2	10	123.10±4.07			
Diastolic (mmHg)	K1	10	80.00±2.25	0.720		
	K2	10	81.50±3.05			

Characteristics of research subjects

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The results of the Shapiro-Wilk test showed that the pre-test and post-test protein carbonyl data did not deviate significantly from the normal distribution (p>0.05), thus allowing the use of parametric statistical tests (Table 2).

Table 2

Data	Group	Shapiro-Wilk	
2	oroup	n	p-value
Protein Carbonyl (pre-test)	K1	10	0.703
	K2	10	0.395
Protein Carbonyl (post-test)	K1	10	0.472
	K2	10	0.183

Normality test results

In group K1 (placebo), protein carbonyl levels decreased significantly after exercise (p<0.05), likely due to natural recovery mechanisms. In group K2 (roselle flower extract), there was no statistically sig-

nificant decrease in protein carbonyl levels (p>0.05), indicating that supplementation had no additional effect beyond physiological adaptations. Data are presented as mean \pm standard error (Fig. 2, Table 3).



Fig. 2. Research results diagram: group K1 (placebo) and group K2 (roselle flower extract)

Table 3

Results of protein carbonyl levels

Difference Test Method	Group	р
Paired t_test	K1 (pre-test and post-test)	0.001*
	K2 (pre-test and post-test)	0.053

Note. *A significant decrease in protein carbonyl levels was observed in Group K1 (placebo) from pre-test to post-test (p=0.001). However, no significant change was observed in Group K2 (roselle flower extract, p=0.053).

The purpose of this investigation was to ascertain the impact of roselle flower extract supplementation after physical exercise on reducing carbonyl protein levels. The findings of this investigation showed that supplementation of roselle flower extract after physical exercise was not significant in reducing protein carbonyl levels. This was also shown in previous research that administration of 500 mg/day of roselle flower extract for 4 weeks was not able to significantly reduce oxidative stress biomarkers [24]. The cause of the less significant in reducing the levels of carbonyl protein in addition to the samples used being too small, it could be because the dose used was not high enough and the duration of the intervention was not long enough, so this has not had an effect on reducing the levels of carbonyl protein. Although we did not find anti-inflammatory and antioxidant effects of roselle flower extract in our research subjects, several previous studies have shown such effects. In a previous study, polyphenols from roselle flowers were given to patients with metabolic syndrome for 4 weeks. According to the results, the extract showed strong anti-inflammatory activity (decrease in interleukin (IL)-6, IL-1ß and IL-8) and increased antioxidant activity (decrease in 8-isoprostane-F2 α and increase in serum paraoxonase activity) [25]. In our study, the overall content of roselle flower extract has not been proven to be able to reduce carbonyl protein levels, so it is possible that there are special contents such as flavonoids and polyphenols that can optimally provide a real effect in reducing oxidative stress. Future studies should investigate the chronic effects of roselle supplementation at doses exceeding 500 mg, over durations longer than four weeks, to assess potential cumulative antioxidant benefits.

The findings of earlier research have demonstrated that roselle flower extract supplementation can reduce the expression of NF-kB proteins that play a role in the liver's production of different pro-inflammatory proteins, so roselle is effectively able to suppress inflammatory effects [26]. Further investigation is necessary to ascertain the precise impact of supplementing with roselle flower extract following physical activity on protein carbonyl reduction. In addition to potentially impairing the function of several proteins, protein oxidation is extremely harmful because it alters the redox balance of cells, disrupts the cell cycle, which eventually leads to the death of neurons [27].

Among the oxidative stress processes, protein carbonyls emerge as biomarkers that cause obvious damage to cells [28]. Carbonylated proteins are normally degraded by the proteasome system; however, excessive accumulation can lead to the formation of protein aggregates, which are resistant to degradation and can cause cellular dysfunction [27]. If the cell's carbonylated proteins build up, they will have an impact and cause cytotoxic effects [27]. In addition protein carbonyls make cells more susceptible to apoptotic triggers in addition to causing structural and functional alterations [29]. As a result, protein carbonyls have gained significance as important markers of oxidative stress, due to their sensitivity as well as their structural and functional ramifications [30]. According to the findings of earlier research, giving rats with diabetes mellitus roselle flower extract significantly lowers their protein carbonyl levels [31]. Although this study did not show a significant reduction in protein carbonyl levels, previous studies have shown that roselle flower extract has antioxidant properties that may help reduce oxidative stress [32]. Roselle antioxidant qualities are widely known [33].

According to other study findings, red and white roselle include a variety of phytochemical components that can prevent the negative effects of oxidative stress, such as organic acids, phenolic acids, anthocyanins, and flavonoids [34]. Roselle flower extract has been shown to reduce oxidative stress in obese rats with myocardial infarction, potentially by increasing superoxide dismutase (SOD) activity, increasing glutathione (GSH) levels, and inhibiting NOX2-mediated ROS production [35]. This is because anthocyanins, phenolic acids, and flavonoids can scavenge free radicals such reactive nitrogen species (RNS) and ROS, hence preventing oxidative stress [36]. So although this review did not significantly reduce protein carbonyl levels, the antioxidant effect caused in the previous study can clearly inhibit oxidative stress. The insignificant decrease in protein levels caused in this study could be impacted by a number of variables, including the quantity of samples and the physiological response of the body of each sample in responding to the pharmacological effects of roselle flower extract consumed.

In the control group given placebo in this investigation, it was demonstrated that there was a noteworthy decrease in protein carbonyl levels 60 minutes after physical exercise. This needs to be further explored that the post-test was conducted 60 minutes after exercise and placebo administration. There are indications that this period of time is the body's physiological response in the mechanism of increasing the body's antioxidant levels such as SOD, CAT, GPx. We know that during physical exercise, the body's metabolic rate increases, which can lead to much higher consumption of oxygen, which raises the generation of reactive oxygen species (ROS) [37].

SOD participates in the disproportionation of superoxide free radicals inside and outside cells so that cell membranes and DNA are protected from

damage by oxygen free radicals [38]. SOD enzyme (SOD) plays an important role in balancing the concentration of biological oxidants, which can protect all aerobic organisms from oxidative damage caused by biological oxidants. SOD is one of the most powerful antioxidant enzymes, which can catalyze O₂to form H₂O₂ to regulate the body's signal transduction [39]. At low concentrations, H₂O₂ acts as a second messenger in redox signaling pathways, modulating inflammation, angiogenesis, and gene expression. However, excessive accumulation of H₂O₂ can contribute to oxidative stress and cell damage [39]. Additionally, H₂O₂ diffuses across the cell membrane through aquaporin aqueous channels, transposing the redox signal from the location where the redox signal is generated to the desired location [40]. So with the duration of the post-test collection in our study there was a range of one hour, at this time the SOD levels increased which will have an impact on reducing oxidative stress triggered by exercise. This is reinforced by previous research that high intensity interval training has been shown to increase SOD levels immediately after the intervention and 3 hours after the intervention also still provides an increase in SOD levels [41].

Research shows that exhausting and prolonged exercise causes inflammation and oxidative stress are harmful to health [42]. The results of previous studies showed that participants who participated in the 50 km ultamaraton running race were shown to provide an increase in protein carbonyl production significantly, the duration and intensity of exercise had an effect on the level of ROS [15]. Another study looked at how running affected the oxidative modification of nucleic acids and discovered that running a marathon instantly caused an acute inflammatory response. This suggests that marathon running also affects endogenous antioxidant activity as an exercise adaptation process of elevated ROS [43].

Another research revealed that 10 weeks of aerobic exercise on a treadmill was shown to reduce protein carbonyl levels [44]. This may be due to an adaptation process that prevents protein carbonyl accumulation during physical exercise. Conversely, it has been demonstrated that a single high-intensity exercise session can raise the lipid peroxidation index [36]. The high intensity of exercise greatly impacts the level of lipid peroxidation that occurs. This is also supported by research from (Ji et al., 2016) that performing a single session of the rate of lipid peroxidation in trained rats' cardiac muscle increased with resistance training [37]. There are inconsistencies in the results of the research that occurred. It should be noted that many factors affect the occurrence of oxidative stress during exercise including the length of intervention, exercise intensity, exercise duration, and data collection period also affect the different results of each study. Acute physical exercise or one session of strenuous exercise is closely associated with an increase in free radical levels, while long-term or chronic exercise of four weeks, eight weeks, or longer is linked to adaptability process that results in an increase in antioxidants in the body [38].

The results of the study prove that exercise can improve mitochondrial function, which is a cell organelle that functions to produce ATP or what is known as an energy warehouse [46]. The reaction mechanism of antioxidant enzymes is very complex and involves several reaction stages and involves various cell organelles. Thus, it can be said that antioxidants in biological systems stop the increase of ROS. Antioxidants prevent the increase in oxidative stress, which reduces oxidative damage to biological components [47]. In fighting free radicals, the enzyme catalase is very important. Since one molecule of catalase can convert millions of H₂O₂ molecules into H₂O and oxygen per second, it has been proven that catalase is one of the most catalytic and efficient enzymes. Due to its outstanding catalytic rate, this enzyme plays an important role in oxidative damage caused by exercise [48]. So in detail, exercise is proven to increase ROS. With the presence of ROS as a trigger for oxidative stress, the body naturally increases the antioxidant defense system in the body such as SOD. Superoxide dismutase (SOD) catalyzes the dismutation of superoxide anion (O₂⁻) to hydrogen peroxide (H₂O₂), which reduces oxidative damage. H2O2 is then further broken down by catalase (CAT) to water (H₂O) and oxygen (O₂) or by glutathione peroxidase (GPx), which converts H₂O₂ to H₂O using glutathione as a cofactor. So antioxidants are related to each other in an effort to neutralize free radicals triggered by exercise [49].

The findings of earlier research that physical exercise interventions were shown to briefly provide an increase in protein carbonyl levels, but there was a downward trend 60 minutes after the physical exercise intervention and this is a physiological response caused during exercise [50]. It is possible that 60 minutes after exercise, increased SOD activity contributes to the decrease in protein carbonyl levels. However, other antioxidant systems, such as catalase (CAT) and glutathione peroxidase (GPx), may also play a role in reducing oxidative damage. Three isoenzymes make up the antioxidant enzyme SOD. By preserving the body's antioxidant defenses and reducing oxidative stress brought on by an increase in free radicals, SOD is the body's first line of enzymatic defense against free radicals [51]. Regular exercise, on the other hand, can boost the antioxidant system

and lower the body's level of free radicals, enhancing tolerance to oxidative stress and regulating the rate of cell damage [52]. So it can be concluded that the decrease in protein carbonyl levels 60 minutes after physical exercise is a physiological response due to physical exercise by increasing SOD levels which function to reduce free radicals.

CONCLUSION

1. Administration of 500 mg of roselle (Hibiscus sabdariffa) extract after physical exercise did not result in a statistically significant reduction in protein carbonyl levels (p>0.05). However, a non-significant downward trend was observed, suggesting that further studies with a larger sample size and longer intervention duration are warranted.

2. The main limitations of this study include the small sample size (n=20) and the single-session intervention rather than a prolonged supplementation period. Future research should investigate higher doses and prolonged administration of roselle flower extract over multiple weeks, considering additional factors such

1. Powers SK, Deminice R, Ozdemir M, Yoshihara T, Bomkamp MP, Hyatt H. Exercise-induced oxidative stress: Friend or foe? J Sport Heal Sci. 2020;9(5):415-25. doi: https://doi.org/10.1016/j.jshs.2020.04.001

2. Kruk J, Aboul-Enein HY, Kładna A, Bowser JE. Oxidative stress in biological systems and its relation with pathophysiological functions: the effect of physical activity on cellular redox homeostasis. Free Radic Res. 2019;53(5):497-521.

doi: https://doi.org/10.1080/10715762.2019.1612059

3. Kraus WE, et al. Physical Activity, All-Cause and Cardiovascular Mortality, and Cardiovascular Disease. Med Sci Sports Exerc. 2019;51(6):1270-81.

doi: https://doi.org/10.1249/MSS.000000000001939

4. Militello R, Luti S, Gamberi T, Pellegrino A, Modesti A, Modesti PA. Physical Activity and Oxidative Stress in Aging. Antioxidants. 2024;13(5):557. doi: https://doi.org/10.3390/antiox13050557

5. Sun Y, Chen P, Zhai B, Zhang M, Xiang Y, Fang J, et al. The emerging role of ferroptosis in inflammation. Biomed Pharmacother. 2020 Jul;127:110108.

doi: https://doi.org/10.1016/j.biopha.2020.110108

6. Steinbacher P, Eckl P. Impact of oxidative stress on exercising skeletal muscle. Biomolecules. 2015 Apr 10;5(2):356-77.

doi: https://doi.org/10.3390/biom5020356

7. Powers SK, Jackson MJ. Exercise-induced oxidative stress: Cellular mechanisms and impact on muscle force production. Physiol Rev. 2008 Oct;88(4):1243-76. doi: https://doi.org/10.1152/physrev.00031.2007

8. Marciniak A, Brzeszczyńska J, Gwoździński K, Jegier A. Antioxidant capacity and physical exercise. Biol Sport. 2009;26(3):197-213.

doi: https://doi.org/10.5604/20831862.894649

as dietary intake, other oxidative stress biomarkers, and inter-individual variability in exercise response.

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REFERENCES

9. He F, Li J, Liu Z, Chuang CC, Yang W, Zuo L. Redox mechanism of reactive oxygen species in exercise. Front Physiol. 2016 Nov 7;7:486.

doi: https://doi.org/10.3389/fphys.2016.00486

10. Campbell JP, Turner JE. Debunking the myth of exercise-induced immune suppression: Redefining the impact of exercise on immunological health across the lifespan. Front Immunol. 2018 Apr 16;9:648.

doi: https://doi.org/10.3389/fimmu.2018.00648

11. Song YR, Kim JK, Lee HS, Kim SG, Choi EK. Serum levels of protein carbonyl, a marker of oxidative stress, are associated with overhydration, sarcopenia and mortality in hemodialysis patients. BMC Nephrol. 2020 Jul 16;21(1):281.

doi: https://doi.org/10.1186/s12882-020-01937-z

12. Leger T, He B, Azarnoush K, Jouve C, Rigaudiere JP, Joffre F, et al. Dietary EPA increases rat mortality in diabetes mellitus, a phenomenon which is compensated by green tea extract. Antioxidants. 2019 Nov 4;8(11):526. doi: https://doi.org/10.3390/antiox8110526

13. Frijhoff J, Winyard PG, Zarkovic N, Davies SS, Stocker R, Cheng D, et al. Clinical Relevance of Biomarkers of Oxidative Stress. Antioxidants Redox Signal. 2015 Nov 10;23(14):1144-70.

doi: https://doi.org/10.1089/ars.2015.6317

14. Nowak K, Zabczyk M, Natorska J, Zalewski J, Undas A. Elevated plasma protein carbonylation increases the risk of ischemic cerebrovascular events in patients with atrial fibrillation: association with a prothrombotic state. J Thromb Thrombolysis. 2024 Oct;57(7):1206-15.

doi: https://doi.org/10.1007/s11239-024-03003-z

15. Vezzoli A, Dellanoce C, Mrakic-Sposta S, Montorsi M, Moretti S, Tonini A, et al. Oxidative Stress Assessment in Response to Ultraendurance Exercise: Thiols

На умовах ліцензії СС ВУ 4.0



Redox Status and ROS Production according to Duration of a Competitive Race. Oxid Med Cell Longev. 2016;2016:6439037.

doi: https://doi.org/10.1155/2016/6439037

16. Thirupathi A, Wang M, Lin JK, Fekete G, István B, Baker JS, et al. Effect of Different Exercise Modalities on Oxidative Stress: A Systematic Review. Biomed Res Int. 2021 Feb 11;2021:1947928.

doi: https://doi.org/10.1155/2021/1947928

17. Arjmand A, Abedi B, Hosseini SA. The Effect of Resistance Training on Malondialdehyde and Protein Carbonyl Concentration in the Heart Tissue of Rats Exposed to Stanozolol. Pharm Biomed Res. 2020;6(4):261-8. doi: https://doi.org/10.18502/pbr.v6i4.5112

18. Setianingsih S, Nurani LH, Rohman A. Effect of the ethanolic extract of red roselle calyx (Hibiscus sabdariffa L.) on hematocrit, platelets, and erythrocytes in healthy volunteers. Pharmaciana. 2018;8(1):266. doi: https://doi.org/10.12928/pharmaciana.v8i2.8738

19. Jin HM, Dang B, Zhang WG, Zheng WC, Yang XJ. Polyphenol and Anthocyanin Composition and Activity of Highland Barley with Different Colors. Molecules. 2022 May 25;27(11):3411.

doi: https://doi.org/10.3390/molecules27113411

20. Ayubi N, et al. Potential of Polyphenolic Compounds in Rosella Flowers on Reducing Oxidative Stress and Inflammation After Exercise: A Systematic Review. Phys Educ Theory Methodol. 2024;24(1):169-79. doi: https://doi.org/10.17309/tmfv.2024.1.20

21. Sapian S, Ibrahim Mze AA, Jubaidi FF, Mohd Nor NA, Taib IS, Abd Hamid Z, et al. Therapeutic Potential of Hibiscus sabdariffa Linn. in Attenuating Cardiovascular Risk Factors. Pharmaceuticals. 2023 May 29;16(6):807. doi: https://doi.org/10.3390/ph16060807

22. Wibawa JC, Arifin MZ, Herawati L. Ascorbic Acid Drink after Submaximal Physical Activity can Maintain the Superoxide Dismutase Levels in East Java Student Regiment. Indian J Forensic Med Toxicol. 2021;15(3):3383-92. doi: https://doi.org/10.37506/ijfmt.v15i3.15824

23. Aritanoga M, Effendi C, Herawati L. Kopi Arabika-Gayo Menurunkan MDA dan Meningkatkan SOD setelah Latihan Fisik Akut Submaksimal pada Pria Sedenter Gayo-Arabica Coffee Decreases MDA and Increases SOD after Single Bout Submaximal Physical Exercise in Sedentary Men. Jurnal Sumberdaya Hayati. 2019;5(2):58-63.

doi: https://doi.org/10.29244/jsdh.5.2.58-63

24. Asgary S, Soltani R, Zolghadr M, Keshvari M, Sarrafzadegan N. Evaluation of the effects of roselle (Hibiscus sabdariffa L.) on oxidative stress and serum levels of lipids, insulin and hs-CRP in adult patients with metabolic syndrome: A double-blind placebo-controlled clinical trial. J Complement Integr Med. 2016 Jun 1;13(2):175-80. doi: https://doi.org/10.1515/jcim-2015-0030

25. Joven J, March I, Espinel E, Fernández-Arroyo S, Rodríguez-Gallego E, Aragonès G, et al. Hibiscus sabdariffa extract lowers blood pressure and improves endothelial function. Mol Nutr Food Res. 2014 Jun;58(6):1374-8. doi: https://doi.org/10.1002/mnfr.201300774

26. Ujianti I, Sianipar IR, Prijanti AR, Hasan I, Arozal W, Jusuf AA, et al. Effect of Roselle Flower Extract (Hibiscus sabdariffa Linn.) on Reducing Steatosis and Steatohepatitis in Vitamin B12 Deficiency Rat Model. Med. 2023 May 28;59(6):1044.

doi: https://doi.org/10.3390/medicina59061044

27. Akagawa M. Protein carbonylation: molecular mechanisms, biological implications, and analytical approaches. Free Radic Res. 2021 Apr;55(4):307-20. doi: https://doi.org/10.1080/10715762.2020.1851027

28. Yu S, Doycheva DM, Gamdzyk M, Gao Y, Guo Y, Travis ZD, et al. BMS-470539 Attenuates Oxidative Stress and Neuronal Apoptosis via MC1R/cAMP/PKA/Nurr1 Signaling Pathway in a Neonatal Hypoxic-Ischemic Rat Model. Oxid Med Cell Longev. 2022 Jan 31;2022:4054938. doi: https://doi.org/10.1155/2022/4054938

29. Martínez-Orgado J, Martínez-Vega M, Silva L, Romero A, de Hoz-Rivera M, Villa M, et al. Protein Carbonylation as a Biomarker of Oxidative Stress and a Therapeutic Target in Neonatal Brain Damage. Antioxidants. 2023 Oct 10;12(10):1839.

doi: https://doi.org/10.3390/antiox12101839

30. Martini S, Austin T, Aceti A, Faldella G, Corvaglia L. Free radicals and neonatal encephalopathy: mechanisms of injury, biomarkers, and antioxidant treatment perspectives. Pediatr Res. 2020 Apr;87(5):823-33. doi: https://doi.org/10.1038/s41390-019-0639-6

31. Mohamed J, Shing SW, Idris MH, Budin SB, Zainalabidin S. The protective effect of aqueous extracts of roselle (Hibiscus sabdariffa L. UKMR-2) against red blood cell membrane oxidative stress in rats with streptozotocininduced diabetes. Clinics. 2013 Oct;68(10):1358-63. doi: https://doi.org/10.6061/clinics/2013(10)11

32. Herranz-López M, Olivares-Vicente M, Encinar JA, Barrajón-Catalán E, Segura-Carretero A, Joven J, et al. Multi-targeted molecular effects of Hibiscus sabdariffa polyphenols: An opportunity for a global approach to obesity. Nutrients. 2017 Aug 20;9(8):907. doi: https://doi.org/10.3390/nu9080907

33. Da-Costa-Rocha I, Bonnlaender B, Sievers H, Pischel I, Heinrich M. Hibiscus sabdariffa L. – A phytochemical and pharmacological review. Food Chem. 2014 Dec 15;165:424-43.

doi: https://doi.org/10.1016/j.foodchem.2014.05.002

34. Morales-Luna E, Pérez-Ramírez IF, Salgado LM, Castaño-Tostado E, Gómez-Aldapa CA, Reynoso-Camacho R. The main beneficial effect of roselle (Hibiscus sabdariffa) on obesity is not only related to its anthocyanin content. J Sci Food Agric. 2019 Jan 30;99(2):596-605. doi: https://doi.org/10.1002/jsfa.9220

35. Si LY, Ali SAM, Latip J, Fauzi NM, Budin SB, Zainalabidin S. Roselle is cardioprotective in diet-induced obesity rat model with myocardial infarction. Life Sci. 2017 Dec 15;191:157-65.

doi: https://doi.org/10.1016/j.lfs.2017.10.030

36. Mattioli R, Francioso A, Mosca L, Silva P. Anthocyanins: A Comprehensive Review of Their Chemical Properties and Health Effects on Cardiovascular and Neurodegenerative Diseases. Molecules. 2020 Aug 21;25(17):3809. doi: https://doi.org/10.3390/molecules25173809

37. Meng Q. The Impact of Physical Exercise on Oxidative and Nitrosative Stress : Balancing the Benefits and Risks. 2024, p. 1-26.

38. Tikhonov IV, Pliss EM, Borodin LI, Sen' VD. Effect of superoxide dismutase on the oxidation of methyl linoleate in micelles inhibited by nitroxyl radicals. Russ Chem Bull. 2016;65(12):2985-7.

doi: https://doi.org/10.1007/s11172-016-1690-7

39. Zheng M, Liu Y, Zhang G, Yang Z, Xu W, Chen Q. The Applications and Mechanisms of Superoxide Dismutase in Medicine, Food, and Cosmetics. Antioxidants. 2023 Aug 27;12(9):1675.

doi: https://doi.org/10.3390/antiox12091675

40. Bienert GP, Møller AL, Kristiansen KA, Schulz A, Møller IM, Schjoerring JK, et al. Specific aquaporins facilitate the diffusion of hydrogen peroxide across membranes. J Biol Chem. 2007 Jan 12;282(2):1183-92.

doi: https://doi.org/10.1074/jbc.M603761200

41. Lee TT, Li TL, Ko BJ, Chien LH. Effect of Acute High-Intensity Interval Training on Immune Function and Oxidative Stress in Canoe/Kayak Athletes. Biology (Basel). 2023 Aug 18;12(8):1144.

doi: https://doi.org/10.3390/biology12081144

42. Mrakic-Sposta S, Gussoni M, Moretti S, Pratali L, Giardini G, Tacchini P, et al. Effects of mountain ultramarathon running on ROS production and oxidative damage by micro-invasive analytic techniques. PLoS One. 2015 Nov 5;10(11):e0141780.

doi: https://doi.org/10.1371/journal.pone.0141780

43. Larsen EL, Poulsen HE, Michaelsen C, Kjær LK, Lyngbæk M, Andersen ES, et al. Differential time responses in inflammatory and oxidative stress markers after a marathon: An observational study. J Sports Sci. 2020;38(18):2080-91. doi: https://doi.org/10.1080/02640414.2020.1770918

44. Molina MN, Ferder L, Manucha W. Emerging Role of Nitric Oxide and Heat Shock Proteins in Insulin Resistance. Curr Hypertens Rep. 2016 Jan;18(1):1. doi: https://doi.org/10.1007/s11906-015-0615-4

45. Ji LL, Kang C, Zhang Y. Exercise-induced hormesis and skeletal muscle health. Free Radic Biol Med. 2016 Sep;98:113-22.

doi: https://doi.org/10.1016/j.freeradbiomed.2016.02.025

46. Lim AY, Chen YC, Hsu CC, Fu TC, Wang JS. The Effects of Exercise Training on Mitochondrial Function in Cardiovascular Diseases : A Systematic Review and Meta-Analysis. Int J Mol Sci. 2022 Oct 19;23(20):12559. doi: https://doi.org/10.3390/ijms232012559

47. Jomova K, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, Valko M. Several lines of antioxidant defense against oxidative stress: antioxidant enzymes , nanomaterials with multiple enzyme – mimicking activities , and low – molecular – weight antioxidants. Arch Toxicol. 2024 May;98(5):1323-67.

doi: https://doi.org/10.1007/s00204-024-03696-4

48. Anwar S, Alrumaihi F, Sarwar T, Babiker AY, Khan AA, Prabhu SV, et al. Exploring Therapeutic Potential of Catalase : Strategies in Disease Prevention and Management. 2024 Jun 14;14(6):697.

doi: https://doi.org/10.3390/biom14060697

49. Wibawa JC, et al. Increased activity of the catalase enzyme after physical exercise as a signal for reducing hydrogen peroxide (H_2O_2): a systematic review. Fizjoterapia Pol. 2024;5:232-8.

doi: https://doi.org/10.56984/8ZG020C7GDL

50. Bloomer RJ, Davis PG, Consitt LA, Wideman L. Plasma protein carbonyl response to increasing exercise duration in aerobically trained men and women. Int J Sports Med. 2007 Jan;28(1):21-5.

doi: https://doi.org/10.1055/s-2006-924140

51. Poblete Aro CE, Russell Guzmán JA, Soto Muñoz ME, Villegas González BE. Effects of high intensity interval training versus moderate intensity continuous training on the reduction of oxidative stress in type 2 diabetic adult patients: CAT. Medwave. 2015 Aug 13;15(7):e6212.

doi: https://doi.org/10.5867/medwave.2015.07.6212

52. Quan H, Koltai E, Suzuki K, Aguiar AS Jr, Pinho R, Boldogh I, et al. Exercise, redox system and neurodegenerative diseases. Biochim Biophys Acta - Mol Basis Dis. 2020 Oct 1;1866(10):165778.

doi: https://doi.org/10.1016/j.bbadis.2020.165778

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